NOTE

# Synthesis of [2'-13C]Biotin

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### Summary

 $[2'-^{13}C]$ Biotin was synthesized in 51% yield by reaction of the hydrolysis product of biotin, 3,4-diaminotetrahydro-2-thiophen-valerianic acid, with  $[^{13}C]$ phosgene.

**Key words:** Carbon-13; [<sup>13</sup>C]biotin; [<sup>13</sup>C]phosgene; C-C, C-H, C-N spin coupling constants

#### Introduction

Biotin (1) is the most important prosthetic group for carbon-dioxide transferring enzymes<sup>1</sup>. Carbon-13 labelled biotin was required as an NMR-probe to study the mechanism of action at the active site of the transcarboxylase enzyme. The methods of Hofmann et al.<sup>2</sup> and Melville et al.<sup>3</sup> were improved to provide yields over 50% compared with 12% by optimising the reaction conditions.

The shortest route to  $[2'^{-13}C]$ biotin  $(\underline{3})$  is synthesis via hydrolysis of biotin to the diamino carboxylic acid  $(\underline{2})$  which is reacted with  $[^{13}C]$ phosgene. Hydrolysis of  $(\underline{1})$  in a glass autoclave at  $160^\circ$  for 4 days and recrystallisation of the resulting product gave 89% yield of  $(\underline{2})$ , but only 65% using the procedure of Hofmann et al.<sup>2</sup>.  $[^{13}C]$ Phosgene was synthesized by modification of published procedures  $^4$ ,  $^5$ .

978 W. A. Etzel and S. Berger

 $[2'^{-13}\text{C}]$ Biotin  $(\underline{3})$  was prepared in 51% yield by slowly warming up a flask with the frozen alkaline solution of diamino acid  $(\underline{2})$  and the condensed  $[^{13}\text{C}]$ phosgene followed by stirring overnight. The isotopic purity was determined by mass spectra and integration of inverse gated decoupled  $^{13}\text{C-NMR}$  spectra  $^6$  to be 91.9%. In addition 1.6 g of unreacted diamino acid  $(\underline{2})$  and 0.8 g of another diastereomer of biotin were isolated.

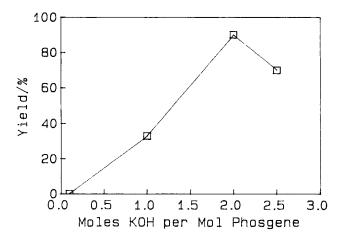


Figure 1. Dependence of the yield on the molar ratio of base to phosgene.

Overall yields were improved because of longer reaction times for the hydrolysation. Only a stoichiometric amount of base was used in the phosgenation reaction compared with nearly 3 equivalents as described by Melville et al.  $^3$  relative to the expected amount of HCl liberated by the reaction of phosgene. Model studies on the reaction of the diamino acid ( $^2$ ) with excess phosgene using various amounts of base showed the dependence of the yield on the molar ratio of the base to the amount of phosgene used (see figure 1).

In the proton-NMR-spectrum the labelling was observed by the change of the AB-systems of the 3-H and the 4-H multiplets to the more complicated ABX-systems (see figure 2).

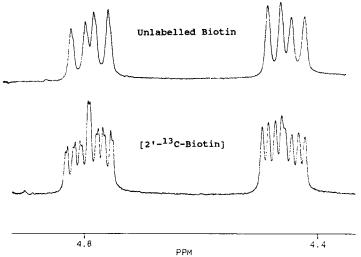


Figure 2. Proton-NMR-spectra of H-3 and H-4 of unlabelled ( $\frac{1}{2}$ ) and [2'- $^{13}$ C]biotin (3).

 $^3\mathrm{J}_{\mathrm{C-H}}$ -couplings were determined by single frequency decoupled  $^{13}\mathrm{C-S}$  spectra:  $^3\mathrm{J}_{\mathrm{C-2'-H-3}}=2.3$  Hz,  $^3\mathrm{J}_{\mathrm{C-2'-H-4}}=1.7$  Hz. The  $^4\mathrm{J}_{\mathrm{C-H}}$ -couplings could not be resolved. The  $^3\mathrm{J}_{\mathrm{C-C}}$ -couplings were determined by normal  $^1\mathrm{H}$ -decoupled  $^{13}\mathrm{C}$ -spectra:  $^3\mathrm{J}_{\mathrm{C-2'-C-3}}=3.9$  Hz,  $^3\mathrm{J}_{\mathrm{C-2'-C-4}}=3.9$  Hz. Two  $^4\mathrm{J}_{\mathrm{C-C}}$ -couplings were observed:  $^4\mathrm{J}_{\mathrm{C-2'-C-2}}=0.9$  Hz,  $^4\mathrm{J}_{\mathrm{C-2'-C-2}}=0.9$  Hz,  $^4\mathrm{J}_{\mathrm{C-2'-C-2}}=0.7$  Hz. The  $^1\mathrm{J}_{\mathrm{C-N}}$ -coupling constants were determined by an inverse  $^{15}\mathrm{N-1}$ H-correlation spectrum  $^7$ :  $^1\mathrm{J}_{\mathrm{C-2'-N-1'}}=^1\mathrm{J}_{\mathrm{C-2'-N-3'}}=18.5$  Hz.  $^1\mathrm{J}_{\mathrm{N-H}}$ -couplings were determined from this spectrum to 94 Hz. The  $^1\mathrm{H}$  and  $^{13}\mathrm{C}$ -signals for the diamino acid sulfate and the diamino acid were assigned by COSY-spectra and inverse C,H-correlation spectra.

### Experimental

#### Materials

[<sup>13</sup>C]Carbon monoxide was purchased from MSD-Isotopes, Montreal, Canada. Biotin was a gift from BASF, Ludwigshafen, Germany. NMR-spectra were recorded on a Bruker AM-400 (400 MHz) or a Bruker AC-300 (300 MHz) spectrometer. Electron-ionization (EI)-mass spectra were obtained on a Varian CH7 mass spectrometer.

980 W. A. Etzel and S. Berger

### 3,4-Diaminotetrahydro-2-thiophenvalerianic acid (2)

A mixture of 8 g ( $\underline{1}$ ) with 100 g barium hydroxide in 500 ml water was placed in a glass autoclave and heated to 160°C for 4 days. The resulting mixture was bubbled with carbon dioxide until neutralization. After filtration the solution was concentrated in vacuo. A yellow-brown oil was obtained which was dissolved in 50 ml of water. Acidification with 2N  $\rm H_2SO_4$  to pH 3.5 yielded a small amount of precipitate which was filtered off. Concentration of the solution to 5 ml and addition of 3-4 ml methanol gave on crystallisation 9.2 g of ( $\underline{2}$ ) sulfate (colorless crystals, mp. 258°-260° C).

 $^{1}$ H-NMR(D<sub>2</sub>O, internal dioxan (=3.53 ppm), pH 4.0): = 4.06-3.96(m, 2H, H-3, H-4), 3.69-3.57(m, 1H, H-2), 3.20-3.13 (dd, 1H, H-5<sub>B</sub>), 2.82(t, 1H, H-5<sub>A</sub>), 2.18(t, 2H, H-9), 1.74-1.14(m, 6H, H-6,H-7,H-8).

 $^{13}$ C-NMR(D<sub>2</sub>O, internal dioxan (=66.6 ppm), pH 4.0): =178.7(C-10), 56.1(C-3), 53.8(C-4), 46.5(C-2), 33.4(C-9), 29.2(C-5), 27.7, 27.0, 23.7(C-6,C-7,C-8).

Neutral diamino carboxylic acid ( $\underline{2}$ ) was prepared by dissolving the diamino acid sulfate in 20 ml of water and titrating this solution with 2N Ba(OH) $_2$  to pH 7.0. The resulting precipitate was filtered off and the solution was concentrated in vacuo to afford 6.3 g of ( $\underline{2}$ ) (slightly yellow crystals, mp. 190°C).

 $^{1}\text{H-NMR}(D_{2}\text{O}, \text{ internal dioxan}(=3.53 \text{ ppm}), \text{ pH } 7.0) : = 3.62-3.42 \text{ (m,} \\ 2\text{H, H-2, H-4}), 3.38(\text{t, 1H, J=3.9 Hz, H-3}), 2.92(\text{dd., 1H, H-5,} \\ 2.55(\text{t, 1H, J=10.7 Hz, H-5}), 1.96(\text{t, 2H, J=7.3 Hz, H-9}), 1.62-1.07(\text{m, 6H, H-6,H-7,H-8}).$ 

 $^{13}$ C-NMR(D<sub>2</sub>O, internal dioxan(=66.6 ppm), pH 7.0):=184.4(C-10), 57.5(C-3), 56.3(C-4), 50.1(C-2), 38.1(C-9), 30.5(C-5), 30.4, 28.4, 26.4 (C-6, C-7, C-8).

[2'-'\*C|Biotin 981

## [13C]Phosgene

The apparatus used for the synthesis of  $^{[13}$ C]phosgene and  $^{[13}$ C]biotin is shown in figure 3.

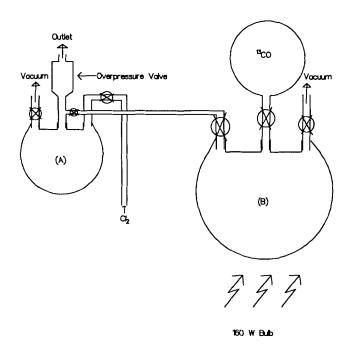


Figure 3. Apparatus for the preparation of  $^{13}$ C-labelled phosgene and reaction with the diamino carboxylic acid ( $\underline{2}$ ).

Chlorine was measured by filling the evacuated three necked flask  $\underline{A}$  until bubbling of the overpressure valve, and was condensed into the evacuated, liquid nitrogen cooled three necked flask ( $\underline{B}$ ). [13C]Carbon Monoxide from flask ( $\underline{C}$ ) was condensed into flask ( $\underline{B}$ ). Warming of flask ( $\underline{B}$ ) with parallel irradiation of the flask with the light of a 160 W bulb and continuing the irradiation 3 hours yielded quantitatively [13C]phosgene.

982 W. A. Etzel and S. Berger

## $[2'-^{13}C]$ Biotin (3)

(1988).

A solution of 4.7 g diamino acid ( $\underline{2}$ ) (0.021 mol) and 6.0 g KOH (0.107 mol) in 50 ml of water was placed in flask ( $\underline{A}$ ). This solution was cooled with liquid nitrogen and the flask was evacuated. The labelled phospene was condensed from flask ( $\underline{B}$ ) to flask ( $\underline{A}$ ). After evacuation of flask ( $\underline{A}$ ) the solution was slowly warmed to room temperature and stirred overnight. Precipitation of labelled biotin commenced after 0.5 h.. Filtration of the solution followed by concentration to 10 ml gave 2.7 g (51.1 % yield) of [2'-13C]biotin, mp. 232°C.

#### REFERENCES

- (1) a. Wood HG and Barden RE Ann. Rev. Biochem. 46:385 (1977).b. Tipton PA and Cleland WW J. Am. Chem. Soc. 110:5866
  - c. Attwood PV and Cleland WW Biochemistry 25:8191 (1986).
  - d. Hansen DE and Knowles JR J. Am. Chem. Soc.  $\underline{107}$ :8304 (1985).
- (2) Hofmann K, Melville DB and Du Vigneaud V J. Biol. Chem.

  141:207 (1941).
- (3) Melville DB, Pierce JG and Partridge CWH J. Biol. Chem. 180:299 (1949).
- (4) Huston JL and Norris TH J. Am. Chem. Soc. 70:1968 (1948).
- (5) Calvin M, Heidelberger C, Reid JC, Tolbert BM and Yankwich
  PE Isotopic Carbon, Wiley, New York, 1949, p. 155.
- (6) Kalinowski HO, Braun S and Berger S Carbon-13 NMR Spectroscopy, Wiley, New York, 1988, p. 49-51.
- (7) Lerner L and Bax A J. Magn. Reson. 69:375 (1986).